

LEVELS OF NUCLEIC ACIDS AND SOME RELATED ENZYMES  
IN SEEDLING OF BETA VULGARIS AS INFLUENCED BY  
SOME PLANT GROWTH REGULATORS

BY

Y.A.H. Mohamed, B.A. Abdel-Ghaffar and M.A.H. El-Beheiry  
Department of Botany, Faculty of Science , Tanta Univer-  
sity, Tanta, A.R. Egypt

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ABSTRACT

The changes in DNA and RNA and some related enzymes of Beta vulgaris L. seedling were studied after treatment with GA<sub>3</sub>, IAA, GA<sub>3</sub>+IAA or HU (hydroxy urea). Seedlings were grown in 24 h dark or in 12 h light+ 12 h dark before hormonal application . GA<sub>3</sub>, IAA and their mixture induced an increase in the DNA and RNA content of seedlings grown in alternate light and dark, and DNA of etiolated seedlings, with an increase in the volume activities of endonuclease and nuclease in etiolated or light-dark- grown seedlings. GA<sub>3</sub> and IAA caused a decrease of RNA content and ribonuclease T<sub>1</sub> activity of etiolated seedlings . The opposite was true with their mixture application. In etiolated seedlings, HU caused an increase in DNA and RNA content with a stimulation in their enzyme activities at the beginning of seedling growth , than a markedly decrease was observed in DNA and RNA accompanied with inhibitory effect of their enzyme activities. The decrease was directly proportion with the progress of age.

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### INTRODUCTION

GA<sub>3</sub> and IAA Have widely many overlapping functions [11,12,13,24,23,28] ; They affect nucleic acids (1,2,4,6,9,28), while hydroxy urea (HU) has an inhibitory effect [16,20,17 25]. This paper deals with studying the change in nucleic acids and three related enzymes (endonuclease, nuclease and ribonuclease T<sub>1</sub> ) during the early growth of sugar beet seedlings as a result of treatment with GA<sub>3</sub> , IAA, GA<sub>3</sub>+IAA or HU .

### MATERIALS AND METHODS

Twenty-five seeded clusters of sugar beet (Beta vulgaris L.) were germinated on moistened cotton in petri dishes at 20°C. Treatment with IAA, GA<sub>3</sub>, mixture of GA<sub>3</sub>+IAA or HU (10<sup>-5</sup> M) was carried out after the appearance of the radicle. Samples were taken at 4-day intervals after the growth regulator treatment. Illumination took place by means of a fluorescent white light (Phytor C.R.H. Hg tube), intensity 1,000 Lux.

#### Extraction of nucleic acids:

Extraction was carried out by a method cited by Marmur [15] and [17] , in which nucleic acids were extracted in Tris-EDTA buffer (pH 8.0) + 1 % SDS + 1 SSC and the lipids were removed by chloroform/isoamyl alcohol and centrifugation took place at 3,000 rpm. The nucleic acids were in

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the supernatant.

DNA determination:

DNA content was determined using the method adopted by Dische and Schwarz [3] by measuring the optical density at 595 nm of the reaction solution resulting from DNA extract and diphenylamine.

RNA determination :

The content was determined using the method adopted by Schneider [22] by measuring the optical density of the reaction solution resulting from RNA extract and orcin.

Extraction of crude enzymes:

Five grams of seedlings were ground in a porcelain mortar with the enzyme buffer, centrifuged to 5,000 rpm and the clear supernatant was taken.

1. Endonuclease activity:

The activity was measured according to the method of Linn and Lehman [14]. The sample was mixed with DNA buffer (pH 8.0). After incubation and centrifugation at 3,000 rpm, an aliquot of 0.1 ml was mixed with 2.99 ml distilled water and measured at 260 nm .

2. Nuclease activity :

According to the method described by Heins et al. [7]

The sample was mixed with borate buffer (pH 8.8) and DNA solution (0.25 % ). After centrifugation at 3,000 rpm, the optical density of the supernatant was measured at 260 nm.

### 3. Ribonuclease T<sub>1</sub> activity :

The activity was measured by Egami et al. [5]. The sample was mixed with Tris-buffer (pH 7.5) and EDTA. After completion of the reaction, centrifugation for 5 min at 3,000 rpm was carried out and the optical density of the supernatant was measured at 260 nm .

## RESULTS

The nucleic acid (DNA and RNA) content of 24 h dark-grown etiolated seedlings of B. vulgaris and of those exposed to 12 h light and 12 h dark are presented in Table 1. It can be observed that application of GA<sub>3</sub>, IAA and their mixture had resulted in a general increase in DNA content of seedlings grown either in 24 h dark or in alternated with light and dark. However, GA<sub>3</sub> proved to be the best DNA stimulator in etiolated seedlings, and IAA in seedlings grown under alternated light and dark periods.

Although GA<sub>3</sub> and IAA showed an inhibition in RNA content, the opposite was true with their mixture in etiolated seedlings . At the same time , GA<sub>3</sub>, IAA and their mixture resulted in an increase in RNA content of seedlings

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grown with alternated light and dark.

Detection of endonuclease, nuclease and ribonuclease  $T_1$  in 4-, 8-, 12- and 16-day-old seedlings of B. vulgaris treated with  $GA_3$ , IAA,  $GA_3 + IAA$  or HU and grown under 24 h dark or alternated light and dark are shown in Table 2. Endonuclease activity was stimulated by  $GA_3$  or IAA in 4-, 8- and 12-day-old etiolated seedlings or in 8-, 12- and 16-day-old of alternated light and dark periods. Although the mixture of  $GA_3$  and IAA induced a stimulatory effect on seedlings grown under alternated light-dark periods, it caused an inhibition in etiolated seedlings. Nuclease activity was stimulated by  $GA_3$ , IAA or their mixture in etiolated or in seedlings with light-dark periods at first three ages, but was inhibited at the last age.

With respect to ribonuclease  $T_1$ , although  $GA_3$  as well as IAA had decreased its activity in the first three ages of all seedlings, a combination of them led to antagonize the harmful effect in etiolated seedlings. Nevertheless,  $GA_3$ , IAA and their mixture had stimulated volume activities of ribonuclease  $T_1$  in seedlings grown in alternate light and dark at the first three ages, yet the amount tended to equalize the control values in 16-day-old seedlings. In seedlings grown with light-dark periods, the volume activities of the three enzymes were inhibited by  $GA_3$  application

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at the last age (16 days). At the same time, DNA contents were decreased.

On the other hand, HU had induced an increase in the volume activities of the three enzymes in 4-day-old etiolated seedling, and both DNA and RNA were increased . Ribonuclease T<sub>1</sub> of the light-dark grown seedlings only was stimulated at the same age and this was accompanied with an increase in RNA contents . A marked inhibition was observed at the last three ages, i.e. the inhibitory effect was directly proportional with the progress in age.

#### DISCUSSION

Growth regulatory may now be considered in the context of what we know are needed to induce growth. Application of the growth regulators took place after the emergence of the radicle, where this time was recorded as zero time. After 16 days, the seedlings cannot continue due to the lack of nutrients necessary to continue their life. Kende and Gardner [8] suggested that auxin might be active via a direct interaction with the genome. In this study , it was found DNA and RNA amounts were increased in seedlings. This suggests that growth regulators tend to increase the activity of the genome by increasing its amount. Degani et al. [2] found that application of GA<sub>3</sub> increased DNA in Cucumis sativus. Kende and Gardner [8] were not able to demonstrate

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promotion of RNA synthesis by IAA as had been observed in this work. Some authors have reported an effect on RNA synthesis and their enzymes [4,8] . The three nucleic acid enzymes (endonuclease, nuclease and ribonuclease  $T_1$  ) were measured as activities in the seedling stage. These activities were stimulated when treated with  $GA_3$ , IAA and  $GA_3 + IAA$ . This suggests that the mechanism of nucleic acid synthesis or degradation was shared or influenced with the plant growth regulators tested. Degani et al. [2] showed that nucleic acids did show a change in etiolated plants, but the degree of variation was lesser than that extracted from plants grown with alternated light and dark. Mohamed [16] showed that etiolation had increased DNA in Sinapis alba and Pisum sativum. Also, several authors [18,17,26,27] confirmed the fact that DNA was stimulated in dark. This means that light or dark was another factor affecting nucleic acid formation or degradation. In confirming that, Osman and Mohamed [20] found that DNA was dark-dependent process and cannot be induced by light.

HU caused an inhibitory effect in the volume activities of the three enzymes of etiolated seedling at the last three ages and in the light-dark grown seedling at all ages. This was accompanied by a decrease in DNA and RNA content.  $GA_3$  caused the same effect in light-dark-grown seedling at the age . These results suggest an existing equilibrium

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between nucleic acids and their enzymes and that growth regulators would affect such equilibrium.

#### REFERENCES

- 1-DAVES, P.J. and GALSTON, A.W. Labeled indole-macromolecular conjugates from growing stem supplied with labeled indole acetic acid. *Plant Physiol.* 47, 435 - 441 (1971).
- 2-DEGANI, Y., ATSMON, D. and HALEVY, A.H. DNA synthesis and hormone-induced elongation in the cucumber hypocotyl. *Nature (London)* 228, 554 - 555 (1970).
- 3-DISCHE, Z. and SCHWARZ, K. *Microchim. Acta* 2,13 (1937).  
 Cited in : STAHL, E. (ed.): *Thin-Layer Chromatography*. Springer Verlag, Berlin (1978).
- 4-EL-BEHEIRRY, M. A. H. Studies on the growth and development of Beta vulgaris L. and Brassica rapa L.M. Sc. thesis, Tanta Univ., A.R. Egypt (1985).
- 5-EGAMI, F., TAKAHASHI, K. and UCHIDA, T. In : Davidson, J.N. and COHEN, W.E. (eds.) : *Progress in Nucleic Acid Research and Molecular Biology* 3, 59-67. Academic Press. New York (1964).
- 6-FAN, D.F. and MAXLACHLAN, G.A. Massive synthesis of RNA and cellulase in the pea epicotyl in response IAA with and without concurrent cell division. *Plant Physiol.* 42, 114 (1967).
- 7-HEINS, J.N., TANIUCHI, H. and AFINSEN, C.B. in : CANLONI,



Delta J. Sci. 12 (1)1988

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- J. and DAVIFS, D. (eds.) : Procedures in Nucleic Acid Research, 79 - 85. Harper and Row, New York (1966).
- 8 - KENDE, H. and GARDNER, G. Hormone binding in plants. Ann. Rev. Plant Physiol. 27, 267 - 290 (1976).
- 9 - KEY, J.L. Hormones and nucleic acid metabolism. Ann. Rev. Plant Physiol. 20, 449 - 474 (1969).
- 10- LANG, A. and NITSAN, J. Relations among cell growth, DNA synthesis and gibberellin action. Ann. New York Acad. Sci. 144, 180 - 190 (1967).
- 11- LEOPOLD, A.C. Antagonism of some gibberellin actions by a substituted pyrimidine. Plant Physiol. 48, 537 - 540 (1971).
- 12- LEOPOLD, A.C. Ethylene as a plant hormone. In Kaldeway, H. and VARDAR, Y. (eds.) : Hormone Regulation in Plant Growth and Development , 245 - 262. Verla Chemie, Weinheim (1972).
- 13- LEVITT, J. Responses of Plants to Environmental Stresses. Academic Press, New York (1972).
- 14- LINN, S. and LEHMAN, I.R. An endonuclease from Neurospora crassa specific for polynucleotides lacking an ordered structure. J. Biol. 240, 1287 - 1293 (1965).
- 15- MARMUR, J. A procedure for the isolation of DNA from microorganisms. J. Mol. Chem. 3, 208 - 218 (1961).

- 16- MOHAMED , Y.A.H. Änderung des DNA-Gehalts in sich streckenden Pflanzengeweben von Sinapis alba und Pisum sativum. Diss. Heidelberg Uni., West Germany (1978).
- 17- MOHAMED, Y.A.H., and CAPERSIUS, I. Wirkung von Gibberellinsäure und FdUrd auf die Menge und die Zusammensetzung der DNA während des Streckungswachstum von Pisum sativum. Z. Pflanzenphysiol. 98, 15 - 23 (1980).
- 18- MOHAMED, Y.A.H., and BOPP, M. Distribution of ploidy in elongated and non-elongated shoot axis of Pisum sativum. Z. Pflanzenphysiol. 98, 25 - 33 (1980).
- 19- MOHAMED , Y.A.H., ABDEL-GHAFFAR, B.A. and EL-BEHEIRY, M.A.H. The growth, pigmentation and carbohydrate content of Beta vulgaris affected by GA<sub>3</sub>, TAA or combination of both and hydroxyurea. Mansoura Univ. Conf. of Agric. Sci. on Food Deficiency Overcoming through Autonomous Efforts in Egypt 3, 632 - 637 (1987).
- 20- OSMAN, M.E.H. and MOHAMED, Y.A.H. Effect of light/dark rhythm on the DNA-content of etiolated Pisum sativum seedling. Abstracts 1st Nat. Cong. Biochem. (Cairo), 104 - 105 (1981).
- 21- OSMAN, M.E. and MOHAMED, Y.A.H. Effect of certain inhibitors on the growth, nucleic acids,

Delta J. Sci. 12 (1)1988

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- and soluble protein contents of Fusarium solani. Delta J. Sci. 8(2), 708 - 721 (1984).
- 22- SCHNEIDER, W.C. Colorimetric analysis of sugars. Methods in Enzymol. XII(A), 557 (1957).
- 23- SKOOG, F. Substances involved in normal growth and differentiation in plants. Brookhaven Symp. Biol. 7, 1- 23 (1954).
- 24- SILBERGER, J. and SKOOG, F. Changes induced by indoleacetic acid in nucleic acid contents of tobacco pith tissue. Science 118, 443 - 444 (1953).
- 25- SINCLAIR, W.K. Hydroxyurea : Differential lethal effects on cultural mammalian cells during the cell cycle. Science 150, 1729 - 1931 (1965).
- 26- VAN OOSTVELDT, P. and VAN PARIJS, R. Effect of light on nucleic acid synthesis and polyploidy level in elongating epicotyl cell of Pisum sativum, Plants (Berl.) 124, 287 - 295 (1975).
- 27- VAN OOSTVELDT, P. and GOETHEM, G. and VAN PARIJS, R. Effect of light on cell elongation, nucleic acid and protein synthesis in hypocotyls of Lupinus angustifolius. Planta (Berl.) 129, 259 - 263 (1976).
- 28- WAREING, P.F. and PHILLIPS, I.D.J. The Control of Growth and Differentiation in Plants. Pergamon Press, Oxford (1973).

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Table 1 : DNA and RNA contents (mg/g fresh wt) of *B. vulgaris* seedlings grown 24 h in dark or 12 h in light + 12 h in dark and treated or not with  $10^{-5}$  M GA<sub>3</sub>, IAA, GA<sub>3</sub>+IAA and HU.

Treatmer	Seedling grown 24 h in dark				Seedling grown 12 in light+ 12 h in dark			
	4	3	2	16	4	8	12	15
	DNA							
Control	2.7±0.3	2.8±0.3	3.5±0.5	2.2±0.2	3.5±0.2	2.5±0.2	3.2±0.3	2.2±0.2
GA <sub>3</sub>	3.1±0.4	6.0±1.1	5.1±1.2	5.2±0.8	3.3±0.3	4.1±0.3	4.8±0.4	1.1±0.1
IAA	3.5±0.5	3.8±0.7	4.1±0.6	3.7±0.5	5.1±0.2	5.5±0.2	5.2±0.3	2.5±0.1
GA <sub>3</sub> +IAA	3.2±0.3	3.4±0.4	3.7±0.5	3.3±0.3	4.5±0.3	3.0±0.4	4.3±0.4	2.6±0.2
HU	4.7±0.6	1.7±0.2	1.6±0.2	1.4±0.1	3.7±0.2	2.7±0.3	2.6±0.4	2.1±0.2
	RNA							
Control	12.1±2.1	12.5±3.1	7.3±0.9	5.8±0.8	9.5±0.6	8.3±0.7	5.1±0.5	3.7±0.4
GA <sub>3</sub>	10.5±0.9	9.3±1.0	6.3±0.8	5.4±0.7	8.9±0.8	11.9±0.7	5.1±0.4	2.3±0.3
IAA	8.0±0.3	8.4±0.9	5.0±0.7	5.0±0.8	13.6±1.2	14.6±1.3	6.9±0.9	5.4±0.7
GA <sub>3</sub> +IAA	12.2±1.4	12.9±1.6	11.1±1.5	10.3±1.4	3.8±1.1	14.2±1.3	6.9±0.9	3.6±0.3
HU	12.0±1.6	6.7±0.8	3.6±0.5	3.1±0.7	10.6±0.9	7.9±1.1	4.5±0.7	3.4±0.3

± = standard error of the mean.

Table 2 : Volume activities of endonuclease, nuclease and ribonuclease T<sub>1</sub> (unit/ml sample) of *B. vulgaria* seedlings grown 24 h in dark or 12 h in light + 12 h in dark and treated or not with 10<sup>-5</sup> M GA<sub>3</sub>, IAA, GA<sub>3</sub>+IAA and HU.

Treatment	Age(day)	Seedling grown 24 h in dark				Seedling grown 12 h in light + 12 h in dark			
		4	8	12	16	4	8	12	16
		Endonuclease							
Control		960±83	1280±114	1379±121	1402±122	1206±88	1379±121	1424±98	1497±121
GA <sub>3</sub>		1018±101	1332±53	1462±132	1322±132	1168±94	1494±112	1699±124	1379±132
IAA		1254±112	1322±112	1411±112	1379±133	1197±98	1613±157	1930±216	1699±123
GA <sub>3</sub> +IAA		762±72	842±78	1126±111	1040±102	1238±76	1386±155	1840±107	1686±166
HU		973±39	864±75	800±72	678±53	880±78	986±88	854±94	618±75
		Nuclease							
Control		10±0.9	11±1.1	11±1.2	6±0.5	12±1.4	11±1.1	13±1.4	10±0.8
GA <sub>3</sub>		13±1.1	14±1.4	12±0.8	11±0.8	13±0.7	13±0.7	14±2.1	2±0.5
IAA		11±1.0	12±1.1	13±1.3	8±0.5	13±1.3	13±2.1	13±1.7	10±0.7
GA <sub>3</sub> +IAA		13±1.5	10±0.9	12±0.9	7±0.4	14±1.5	13±1.6	12±0.8	9±0.9
HU		14±1.3	6±0.7	8±0.8	5±0.5	12±1.1	11±0.9	10±0.9	8±0.8
		Ribonuclease T <sub>1</sub> (x 1000)							
Control		543±31.7	1122±10	630±5.4	305±7.7	739±11.4	649±3.2	421±1.2	378±1.3
GA <sub>3</sub>		624±5.7	568±5.5	480±4.4	493±5.5	793±3.3	930±2.3	521±3.2	337±4.2
IAA		605±0.5	663±0.7	421±3.4	543±5.7	1064±9.5	1136±3.3	849±5.7	421±3.4
GA <sub>3</sub> +IAA		936±10	865±8.9	961±8.9	421±3.4	805±4.3	1139±9.9	892±9.9	337±3.4
HU		1164±3.9	549±5.7	490±3.4	332±2.5	830±6.7	605±4.8	509±4.4	331±2.1

± = standard error of the mean

## مستوى الاحماض النووية وبعض أنزيماتها فى بيارات

### نبات البنجر تحت تأثير بعض منظمات النمو

د. يوسف عابد حسن محمد - د. بهيه عبد السلام - د. محمد أحمد البحيرى  
قسم النبات - كلية العلوم - جامعة طنطا - طنطا - مصر .

أوضحت النتائج أن معاملة البيارات بحمض الجبريليك أو أندول حمض الخليك أو بمخلوطهما معا قد أدى الى زياده الحمض النوويين فى البيارات المستتبه فى ضوء يعقبه ظلام وصاحب ذلك زياده فى نشاط أنزيماتهما اندونيكلييز ونيكلييز والريبونيكلييز ت<sub>1</sub> ، بينما زاد الحمض النووى (دن أ) فقط فى البيارات المستتبه فى الظلام وصاحب ذلك زياده أنزيم أندونيكلييز ونيكلييز . كما أدت المعاملة بحمض الجبريليك وأندول حمض الخليك كل على حده فى البيارات المستتبه فى الظلام الى نقص فى تركيز الحمض النووى ( رن أ ) وصاحب نقصا فى نشاط أنزيم الريبونيكلييز ت<sub>1</sub> - بينما أدت المعاملة بمخلوطهما معا الى نتائج عكس السابقة أى الى زياده فى تركيز ( رن أ ) صاحبها زياده فى نشاط أنزيم ريبونيكلييز ت<sub>1</sub> .

أحدث أستخدام مادة الهيدروكسى يوريا زياده فى تركيز الحمضين النوويين مع زياده فى نشاط الانزيمات الثلاثه فى بدايه عمر البيارات ( ٤ أيام ) ثم لوحظ نقصا ملحوظا فى تركيز الحمضين النوويين صاحب نقصا فى نشاط أنزيماتهما وأزداد هذا النقص مع التندم فى عمر البيارات .